



# Malondialdehyde Microplate Assay Kit User Manual

**Catalog # ASK1011**

Detection and Quantification of Malondialdehyde (MDA) Content in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

**For research use only. Not for diagnostic or therapeutic procedures.**

Bioworld Technology, Inc. (USA)  
Email: [info@bioworld.com](mailto:info@bioworld.com)  
Web: [www.bioworld.com](http://www.bioworld.com)

Bioworld technology, co. Ltd. (China)  
Email: [info@biogot.com](mailto:info@biogot.com)  
Web: [www.biogot.com](http://www.biogot.com)



I. INTRODUCTION .....2

II. KIT COMPONENTS .....3

III. MATERIALS REQUIRED BUT NOT PROVIDED .....3

IV. SAMPLE PREPARATION .....4

V. ASSAY PROCEDURE ..... 5

VI. CALCULATION ..... 6

VII. TYPICAL DATA.....7



**I. INTRODUCTION**

Quantification of lipid peroxidation is essential to assess oxidative stress in pathophysiological processes. Lipid peroxidation forms Malondialdehyde (MDA) and 4-hydroxynonenal (4-HNE), as natural bi-products. Measuring the end products of lipid peroxidation is one of the most widely accepted assays for oxidative damage. MDA Microplate Assay Kit provides a convenient tool for sensitive detection of the MDA in a variety of samples. The MDA in the sample is reacted with Thiobarbituric Acid (TBA) to generate the MDA-TBA adduct. The MDA-TBA adduct can be easily quantified colorimetrically ( $\lambda = 532 \text{ nm}$ ).

**II. KIT COMPONENTS**

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Reaction Buffer I	10 ml x 1	4 °C
Reaction Buffer II	1 ml x 1	4 °C
Dye Reagent A	Powder x 1	4 °C
Dye Reagent B	5 ml x 1	4 °C
Standard (1 mmol/L)	1 ml x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

**Note:**

**Dye Reagent A:** add 5 ml distilled water and put it in water bath of 70 °C, shake it occasionally to dissolve before use.

**Dye Reagent Working Solution:** when Dye Reagent A is cold, add 5 ml Dye Reagent B, mix.

**III. MATERIALS REQUIRED BUT NOT PROVIDED**

1. Microplate reader to read absorbance at 532 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Ice
7. Centrifuge
8. Timer



**IV. SAMPLE PREPARATION**

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonication 3s, interval 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For serum or plasma samples

Detect directly.



V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Standard	Sample	Blank
Standard	10 µl	--	--
Sample	--	10 µl	--
Assay Buffer	--	--	10 µl
Reaction Buffer I	100 µl	100 µl	100 µl
Mix.			
Reaction Buffer II	10 µl	10 µl	10 µl
Dye Reagent Working Solution	100 µl	100 µl	100 µl
Mix, put the plate into the oven, 90 °C for 30 minutes, when cold, record absorbance measured at 532nm.			

**Note:**

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
  - 2) The concentrations can vary over a wide range depending on the different samples.
- For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.

**VI. CALCULATION**

1. According to the protein concentration of sample

$$\begin{aligned} \text{MDA } (\mu\text{mol/mg}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (V_{\text{Sample}} \times \\ & C_{\text{Protein}}) \\ &= (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / C_{\text{Protein}} \end{aligned}$$

2. According to the weight of sample

$$\begin{aligned} \text{MDA } (\mu\text{mol/g}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (W \times \\ & V_{\text{Sample}} / V_{\text{Assay}}) \\ &= (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / W \end{aligned}$$

3. According to the quantity of cell or bacteria

$$\begin{aligned} \text{MDA } (\mu\text{mol}/10^4) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (N \times \\ & V_{\text{Sample}} / V_{\text{Assay}}) \\ &= (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / N \end{aligned}$$

4. According to the volume of sample

$$\begin{aligned} \text{MDA } (\mu\text{mol/ml}) &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / V_{\text{Sample}} \\ &= (\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) \end{aligned}$$

$C_{\text{Protein}}$ : the protein concentration, mg/ml;

$C_{\text{Standard}}$ : the standard concentration, 1 mmol/L = 1  $\mu\text{mol/ml}$ ;

W: the weight of sample, g;

N: the quantity of cell or bacteria,  $N \times 10^4$ ;

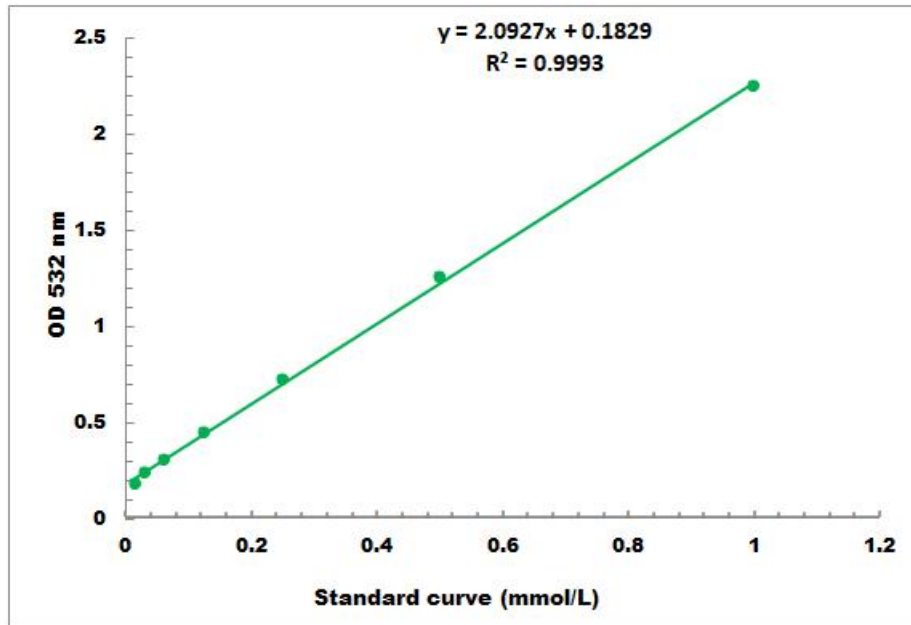
$V_{\text{Sample}}$ : the volume of sample, 0.01 ml;

$V_{\text{Standard}}$ : the volume of standard, 0.01 ml;

$V_{\text{Assay}}$ : the volume of Assay buffer, 1 ml;

**VII. TYPICAL DATA**

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.01 mmol/L - 1 mmol/L